



Antifungal Activities of Five Commercial Extracts Against *Alternaria alternata*

Bachir Raho Ghalem (Corresponding Author)

Laboratory of Process Engineering and Chemistry of Solutions, University of Mascara, Algeria

Email: bachir_rahoo@yahoo.fr

Boudali Talia

Laboratory of Process Engineering and Chemistry of Solutions, University of Mascara, Algeria

Hakem Omar

Laboratory of Process Engineering and Chemistry of Solutions, University of Mascara, Algeria

Article History

Received: May 9, 2020

Revised: June 28, 2020

Accepted: July 30, 2020

Published: August 5, 2020

Copyright © 2020 ARPG
& Author

This work is licensed
under the Creative
Commons Attribution
International



BY: Creative Commons
Attribution License 4.0

Abstract

The inhibitory effects of five commercial extracts (*Allium sativum*, *Chamaemelum nobile*, *Thymus vulgaris*, *Zingiber officinale* and *Ricinus communis*) against *Alternaria alternata* were tested three concentrations (2.5, 5, 12.5 µl/ml) *in vitro*. *T. vulgaris* and *R. communis* extracts both exhibited the most effective antifungal activity against *A. alternata* with diameter of inhibition zones of 54 mm. The *C. nobile* extract exhibited a lower degree of inhibition 24.5 to 45 mm at three concentrations. The antifungal indices of *T. vulgaris* and *R. communis* extracts at three concentrations against *A. alternata* were all 98.14%, while this of *C. nobile* extract was 43.52% at 12.5 µl/ml. The results show that the five commercial extracts have potential for the development of natural antifungal agents, which could be an alternative to chemicals for control of phytopathogenic fungi on fruits or vegetables.

Keywords: Commercial extracts; Antifungal activities; *Alternaria alternata*.

1. Introduction

Plant pathogenic fungi negatively affect a large number of important fruits and vegetables, and limit crop production worldwide, especially in developing countries [1, 2]. The use of chemical control to these pathogens increase the productivity of the crop but it is inappropriate and nondiscriminatory use for human and animal health, as well as contaminating the environment [3, 4]. The search of new fungicides effective, biodegradable and with greater selectivity is necessary to face chemicals' related problems [5]. Within this context is the utilization of plant extracts which are natural sources of antimicrobial substances, regarded as safe and degraded by natural soil microbes; they do not pose any health residual or environmental problems at any concentration which they are used [6, 7]. Antifungal potential of plant extracts has been demonstrated in several works [1, 8, 9]. For example, there are studies evaluating the inhibitory activity of extract on plant pathogenic fungi; **Arora and Kaushik** [10] investigated ginger with 40 different plant extracts for their activity against soybean fungal pathogens as *F. oxysporum* and they published that ginger inhibit their mycelial growth. **Saha, et al.** [11], tested ethanol and aqueous extracts of 30 plants toward the development of ecofriendly antifungal compounds for controlling pathogens responsible fungal diseases of tea (*Pestalotiopsis theae*. (Saw.) Stey., *Colletotrichum camelliae*. Mess., *Curvularia eragrostidis*. (P. Hennings) Meyer, and *Botryodiplodia theobromae*). Results of this study showed that ethanol and aqueous extracts of *Allium sativum*. L., *Datura metel*. L., *Dryopteris filix-mas*. (L.) Schott, *Zingiber officinale*. Rosc., *Smilax zeylanica*. L., *Azadirachta indica*., A. Joss. and *Curcuma longa*. L. recorded 100% inhibition of spore germination. **Al-Rahmah, et al.** [3] evaluated fungal activity of five methanolic plant extracts from *Lantana camara*, *Salvadora persica*, *T. vulgaris*, *Z. officinale* and *Ziziphus spina-christi* on tomato phytopathogenic fungi, *F. oxysporum*, *Pythium aphanidermatum* and *Rhizoctonia solani*. They found that methanolic extracts from *T. vulgaris* and *Z. officinales* were strongly active on these phytopathogenic fungi. **Alemu, et al.** [12] tested antifungal effects of 20 plants against *Colletotrichum gloeosporioides*. They showed that *Datura stramonium* L. methanol extract exhibited a very good antifungal effect on the tested fungus, *Datura stramonium* L. and *E. globulus* Labill. of leaf extracts inhibited the pathogenic spore germination more than the other extracts (14.7% and 15.7%, respectively).

Alternaria alternata is one of the most common saprophytes found throughout the world [13]. *A. alternata* cause a range of diseases with economic impact on a large variety of important agronomic host plants and fruits including potatoes, pomegranate, almond, kiwi, aloe vera, tomato, ginseng, citrus, banana, pepper, water hyacinth, *Lantana camara*, and *Amaranthus* spp. **Dube** [14]. This species has been clinically associated with asthma, allergic rhinosinusitis, hypersensitivity, oculomycosis, onychomycosis, skin infections, and allergic bronchopulmonary mycosis [15]. *Alternaria alternata* is also one of the most important species of *Alternaria* that produces AAL toxins, causing many problems for humans and animals and endangering their health [16]. In addition, Resistance to fungicide has been shown in this species [17-20]. The aim of this work was to evaluate *in vitro* the potential

antifungal activity of five commercial extracts (*Allium sativum*, *Chamaemelum nobile*, *Thymus vulgaris*, *Zingiber officinale* and *Ricinus communis*) against *Alternaria alternata*, in order to verify possible inhibition activity.

2. Materials and Methods

2.1. Commercial Extracts

The following five tested extracts were produced by the Egyptian company El Capitaine (CAPPHARM) for the extraction of oils from natural and cosmetic plants: *Allium sativum*, *Chamaemelum nobile*, *Thymus vulgaris*, *Zingiber officinale* and *Ricinus communis*.

2.2. Fungal Isolates

The fungi *Alternaria alternata* was isolated from lentil seed (Syria R3) and was cultured on to Sabouraud Dextrose Agar at 28° C for 7 days, in experimental farm of nature and life sciences faculty of Mascara University.

2.3. Anti-Fungal Assay

The method of Luan, *et al.* [21] and Hasnaoui, *et al.* [22] with modifications was employed for antifungal evaluation of the selected extracts, which were tested at 2.5 ; 5 et 12.5 µl/ml concentrations against *A. alternata* added to 20 ml of sterilized potato dextrose agar in 9 cm Petri dishes. After the mixture was cooled in the plate (6.0 cm diameter), 5.0 mm diameter of fungi mycelium was transferred to the test plate and incubated at 28 ± 4°C for 3–7 days. When fungi mycelium reached the edges of control plate (without the presence of extracts), the antifungal index was calculated as follows:

$$\text{Antifungal index (\%)} = (1 - D_a/D_b) \times 100$$

where, D_a =the diameter of growth zone in the experimental plate (mm), D_b = the diameter of growth zone in the control plate (mm).

3. Results and Discussion

3.1. Antifungal Activity

The antifungal activity of the selected extracts by direct contact against *A. alternata* was qualitatively assessed by the presence or absence of the inhibition zone. The antifungal activity is summarized in Table 1.

Table-1. Diameter of inhibition zones (mm) of five extracts against *A. alternata* by direct contact method

Extracts	Extract concentrations			
	2.5µl/ml	5µl/ml	12.5µl/ml	control
<i>A. sativum</i>	52	50	47	54
<i>C. nobile</i>	45	42	24.5	54
<i>T. vulgaris</i>	54	54	54	54
<i>Z. officinale</i>	48	42	30	54
<i>R. communis</i>	54	54	54	54

The results revealed that the selected extracts showed a higher antifungal activity with diameter of inhibition zones ranged between 24.5 to 54 mm. Among extracts, *T. vulgaris* and *R. communis* exhibited the most effective antifungal activity with diameter of inhibition zones of 54 mm in the three concentrations of extracts, followed by *A. sativum* (47 to 52 mm), *Z. officinale* (30 to 48 mm) and *C. nobile* (24.5 to 45 mm) which is the weakly active on *A. alternata* in comparison to other extracts.

The antifungal indexes of the five selected extracts against *A. alternata* were shown in Table 2. Results showed that *T. vulgaris* and *R. communis* extract induced 98.14% antifungal index of *A. alternata* with three concentrations (2.5, 5 and 12.5 µl/ml). Similarly, *A. sativum* extract exhibited strong antifungal action with antifungal indices of 94.44 % at concentration of 2.5µl/ml and decrease to 90.74 and 85.18 at concentration of 5 and 12.5µl/ml, respectively.

C. nobile extract which exhibited the lowest antifungal activity induced 81.48 % antifungal index at concentration of 2.5µl/ml and decrease to 75.93 and 43.52 at concentration of 5 and 12.5 µl/ml respectively.

Table-2. Antifungal index of five extracts against *A. alternata* (%)

Extracts	Extract concentrations		
	2.5µl/ml	5µl/ml	12.5µl/ml
<i>A. sativum</i>	94.44	90.74	85.18
<i>C. nobile</i>	81.48	75.93	43.52
<i>T. vulgaris</i>	98.14	98.14	98.14
<i>Z. officinale</i>	87.04	75.93	53.70
<i>R. communis</i>	98.14	98.14	98.14

In this study, extracts of the five plants (*A. sativum*, *C. nobile*, *T. vulgaris*, *Z. officinale* and *R. communis*) were evaluated for their antifungal activities on *A. alternata*. *T. vulgaris* and *R. communis* extract were most active against this phytopathogenic fungi. Thyme oil is one of the 10 most commercial oils worldwide, since it is used as a natural

food preservative, has considerable antioxidant, antibacterial, and antifungal effects, and is used as an aromatic additive to a variety of foods and drinks, as well as in personal care products (perfumes, cosmetics, soaps, oral solutions) [23]. Several works reported the antifungal effects of *Thymus vulgaris* extracts against *A.alternata*. Indeed, Segvić Klarić, *et al.* [24] compare the antifungal activities of essential oil of thyme (*Thymus vulgaris* L.) and pure thymol on different mould species (*Aspergillus*, *Penicillium*, *Alternaria* (include *A.alternata*), *Ulocladium*, *Absidia* and *Mucor*, *Cladosporium*, *Trichoderma* and *Rhizopus*, and *Chaetomium*) isolated from damp dwellings and found that Thymol exhibited approximately three-times stronger inhibition than essential oil of thyme. The MIC and MFC of thyme oil on *A.alternata* were 4.70 µg /ml and 9.40 µg /ml, respectively. In other study, Hadizadeh, *et al.* [25] investigated the antifungal effect of essential oils obtained from some medicinal plants of Iran (*Urtica dioica* L., *Thymus vulgaris* L., *Eucalyptus* spp., *Ruta graveolens* L. and *Achillea millefolium* L.) against *A. alternata*. Both *U. dioica* and *T. vulgaris* oils exhibited antifungal activity against *A. alternata*. The thyme oil exhibited a lower degree of inhibition 68.5 and 74.8% at 1500 and 2000 ppm, respectively. In similarly to Segvić Klarić, *et al.* [24], Perina, *et al.* [26] reported that thyme essential oil from leaves and its major compound thymol had minimum inhibitory concentrations (MIC_s) of 500 and 250 µg mL⁻¹ respectively against *A. alternata*.

In other study carried out by Puškárová, *et al.* [27] on antimicrobial activity of Six essential oils (from oregano, thyme, clove, lavender, clary sage, and arborvitae) against pathogenic (*E. coli*, *S. typhimurium*, *Y. enterocolitica*, *S. aureus*, *L. monocytogenes*, and *E. faecalis*) and environmental bacteria (*B. cereus*, *A. protothormiae*, *P. fragi*) and fungi (*C. globosum*, *P. chrysogenum*, *C. cladosporoides*, *A. alternata*, and *A. fumigatus*). The MIC and MFC of thyme oil on *A.alternata* were 0.025 and 0.05 (% w/v), respectively. Recently, In vitro antifungal activities of vapours of four plant essential oils, cinnamon oil, fennel oil, origanum oil and thyme oil, were investigated by Hong, *et al.* [28] during in vitro conidial germination and mycelial growth of *A. alternata* causing the tomato leaf spots to find eco-friendly alternatives for chemical fungicides. The four plant essential oils showed different antifungal activities against in vitro conidial germination of *A. alternata* in dose-dependent manners. One µl/disc of thyme oil slightly decreased the mycelial growth by ca. 84.3% and increasing to 2 µl/disc led to more decrease in the mycelial growth showing ca. 39.1% compared to that of untreated control. The antimicrobial activities of *Thymus vulgaris* extract is mostly believed to be related to the thymol and carvacrol contents of the oil [29, 30]. Several studies were reported antimicrobial effects of thymol alone or in combination with carvacrol [31-34]. They cause structural and functional disturbances in the cellular membrane [35]. Thymol is lipophilic compound, that alone or with carvacrol, can change the cell membrane fluidity and permeability [36]. In addition to this, the compound can changes the cell membrane in fungi by the affect the function of the cell membrane enzymes that catalyzes the synthesis of the cell wall polysaccharide compounds such as β -glucan and inhibit the growth of cells [37, 38]. *R.communis* extracts exhibited also similar and stronger antifungal effect on *A. alternata* as *T. vulgaris* extracts. Comparing our results with other researchers, they are in good agreement with those of Jassim [39] who tested antifungal activity of *Conocarpus lancifolius*, *Ricinus communis*, *Nerium oleandra* and *Clerodendron inerme* extracts on the fungus *A. alternata* the causal agent of leaves spots of Date Palm. Regarding the treatment of *R. communis* extract 10 and 15% concentrations the inhibition percentage of mycelial growth were 72 and 74.44% respectively. The antifungal activity of *R. communis* extracts is mainly attributed to the presence of camphor [40], which reported to have antimicrobial properties [41, 42].

A. sativum extracts exhibited also remarkable antifungal effects on *A. alternata* near to those of two precedent extracts (*T. vulgaris* and *R. communis*). The results obtained in our screen are in agreement with published results for Taskeen, *et al.* [43], Şesan, *et al.* [44], Alseeni, *et al.* [45] which reported that the extract of *A. sativum* possessed antifungal activity on *A. alternata*. The antimicrobial activity of garlic is believed to be due to the effect of allicin (diallyl thiosulfinate), ajoene, and other sulfite compounds [46].

For *Z. officinale* extracts, it exhibited less antifungal activity than the precedent tested extracts. The results of our study are in line with the report by many researchers [47]; Sharma and Tiwari [48]; Osman, *et al.* [49]; Ahmad and Qureshi [50]; Alseeni, *et al.* [45], which showed the extracts from ginger possess 1antibacterial activity against *A. alternata*. The gingerol and shogaol are the major components of ginger, which 1could be responsible agents for the antimicrobial properties of ginger. They are phenolic compounds causing 1rupture of the bacterial cell membrane and loss of their properties [51] and greater loss of cell contents 1or critical output of molecules and ions can lead to cell death [52, 53].

Comparatively, *A. alternata* were less sensitive to the inhibitory activity of the *C. nobile* extracts than other four tested extracts. To our best knowledge, therefore, this is the first study to determine the antifungal activity of *C. nobile* extract on *A. alternata*. In contrary to our find, [54] tested sixty nine for its antimicrobial activity against three bacteria (*S. aureus*, *S. enteritidis*, *P. aeruginosa*) and three fungi (*A. alternata*, *A. niger*, *P. digitatum*), and among these tested oils, *Anthemis nobilis* (currently *C. nobile*) don't has activity on *A. alternata*. Some studies have been performed concerning the antifungal activity of essential oils or extracts of other *Chamaemelum* species or on *Alternaria* sp. Rizwana, *et al.* [55] have screened *in-vitro* the antimicrobial activity of five organic solvents extracts (Chloroform, ethyl acetate, acetone, ethanol and methanol) of *Matricaria aurea* (golden chamomile) on *S. aureus*, *B. subtilis*, *S.pyogenes*, *E. faecalis*, *E. coli*, *P.aeruginosa*, *K. pneumonia*, *F. oxysporum*, *F. solani*, *A. alternata*, *A. niger*, *A. flavus* and *C. gleosporoides* and found that all extracts inhibit the mycelial growth of *A. alternata* in a percentage ranged between 50.74-100%. In other study, the volatile oil of *Anthemis nobilis* (currently *C. nobile*) inhibited the growth of dermatophytos, *Alternaria* sp., *Aspergillus fumigatus* and *A. parasiticus* [56]. The antimicrobial properties of chamomile have been well documented in some studies [57-59]. Compounds in the essential oil of chamomile were effective against *Staphylococcus* and *Candida* [60]. Of chamomile's essential oil components, α-bisabolol had the strongest activity against Gram-positive and Gram-negative bacteria. Chamazulene

also had strong antimicrobial activity. Spiroethers had weak activity against Gram-positive bacteria but were inactive against Gram-negative bacteria [61]. German chamomile esters and lactones showed activity against *Mycobacterium tuberculosis* and *M. avium* [62]. Chamazulene, α -bisabolol, flavonoids and umbelliferone displayed antifungal properties against *Trichophyton mentagrophytes*, *T. rubrum* and *Candida albicans* [61, 63-65].

4. Conclusion

In this study, we investigated the antifungal activities of five plant extracts (*A. sativum*, *C. nobile*, *T. vulgaris*, *Z. officinale* and *R. communis*) against *A. alternata*. Our study demonstrated that the five plant extracts showed good antifungal activities against this fungi. In particular, *T. vulgaris* and *R. communi* offer effective inhibition activity to the growth of the *A. alternata*. Even at low concentrations, these extracts showed strong antifungal activity. Thus, the results show that the five plant extracts have potential for the development of natural preservatives as alternative to antibiotics and artificial preservatives both of which can be toxic at certain concentrations.

References

- [1] Dellavalle, P. D., Cabrera, A., Alem, D., Larranaga, P., Ferreira, F., and Dalla, R. M., 2011. "Antifungal activity of medicinal plant extracts against phytopathogenic Fungus *Alternaria* spp, Chil." *J. Agric. Res.*, vol. 71, pp. 231-239.
- [2] Shuping, D. S. S. and Eloff, J. N., 2017. "The use of plants to protect plants and food against fungal pathogens: A review." *Afr. J. Tradit. Complement*, vol. 14, pp. 120-127.
- [3] Al-Rahmah, A. N., Mostafa, A. A., Abdel-Megeed, A., Yakout, S. M., and Hussein, S. A., 2013. "Fungicidal activities of certain methanolic plant extracts against tomato phytopathogenic fungi." *African Journal of Microbiology Research*, vol. 7, pp. 517-524.
- [4] Xie, Y., Wang, Z., Huang, Q., and Zhang, D., 2017. "Antifungal activity of several essential oils and major components against wood-rot fungi." *Ind. Crops Prod.*, vol. 108, pp. 278–285.
- [5] Askarne, L., Talibi, I., Boubaker, H., Boudyach, E. H., Msanda, F., Saadi, B. S., and Ait, B. A. A., 2012. "In vitro and in vivo antifungal activity of several Moroccan plants against *Penicillium italicum*, the causal agent of citrus blue mold." *Crop Prot*, vol. 40, pp. 53-58.
- [6] Kim, Y. M., Lee, C. H., Kim, H. G., and Lee, H. S., 2004. "Anthraquinones isolated from *Cassia tora* Leguminosae seed show an antifungal property against phytopathogenic fungi." *J. Agric. Food Chem.*, vol. 52, pp. 6096-6100.
- [7] Yang, X., Ma, X., Yang, L., Yu, D., Qian, Y., and Ni, H., 2010. "Efficacy of *Rheum officinale* liquid formulation on cucumber powdery mildew." *Biol. Cont.*, vol. 522, pp. 167–173.
- [8] Curir, P., Dolci, M., and Galeotti, F., 2005. "Aphytoalexin-like flavonol involved in the carnation *Dianthus caryophyllus*- *Fusarium oxysprum* f. sp. *dianthi* pathosystem." *J. Phytopathol*, vol. 153, pp. 65–67.
- [9] Kumar, A., Shukla, R., Singh, P., Prasad, C. S., and Dubey, N. K., 2008. "Assessment of *Thymus vulgaris* L. essential oil as a safe botanical preservative against post-harvest fungi infestation of food commodities." *Innovat. Food Sci. Emerg. Technol.*, vol. 9, pp. 575–580.
- [10] Arora, C. and Kaushik, R. D., 2003. "Fungicidal activity of plants extracts from Uttaranchal hills against soybean fungal pathogens." *Allelopathy J.*, vol. 112, pp. 217–228.
- [11] Saha, D., Dasgupta, S., and Saha, A., 2005. "Antifungal activity of some plant extracts against fungal pathogen of tea (*Camellia sinensis*)." *Pharma. Biol.*, vol. 43, pp. 87-5.
- [12] Alemu, K., Ayalew, A., and Weldetsadik, K., 2014. "Evaluation of antifungal activity of botanicals for postharvest management of mango anthracnose *Colletotrichum gloeosporioides*." *International Journal of Life Sciences*, vol. 8, pp. 1-6.
- [13] Mohammadi, A. and Bahramikia, S., 2019. "Molecular identification and genetic variation of *Alternaria* species isolated from tomatoes using ITS1 sequencing and inter simple sequence repeat methods." *Curr. Med. Mycol.*, vol. 5, pp. 1-8.
- [14] Dube, J. P., 2014. *Characterization of Alternaria alternata isolates causing brown spot of potatoes in South Africa. Master Thesis in Science (Agriculture) Plant Pathology*. University of Pretoria.
- [15] Mirhendi, H., Fatemi, M. J., Bateni, H., Hajabdolbaghi, M., Geramishoar, M., Ahmadi, B., and Badali, H., 2013. "First case of disseminated phaeohiphomycosis in an immunocompetent individual due to *Alternaria malorum*." *Med. Mycol.*, vol. 51, pp. 196–202.
- [16] Barkai-Golan, R. and Paster, N., 2011. *Mycotoxins in fruits and vegetables*. New York: Elsevier.
- [17] Avenot, H. F. and Michailides, T. J., 2007. "Resistance to boscalid fungicide in *Alternaria alternata* isolates from pistachio in California." *Plant Disease*, vol. 91, pp. 1345-50.
- [18] Dry, I. B., Yuan, K. H., and Hutton, D. G., 2004. "Dicarboximide resistance in field isolates of *Alternaria alternata* is mediated by a mutation in a two-component histidine kinase gene." *Fungal Genetics and Biol.*, vol. 41, pp. 102-8.
- [19] Ozkilinc, H. and Kurt, S., 2017. "Screening fungicide resistance of *Alternaria* pathogens causing *Alternaria* blight of pistachio in Turkey. YYÜ TAR BİL DERG." *YYU. J. AGR. SCI.*, vol. 27, pp. 543-549.
- [20] Yang, He, M. H., Ouyang, H. B., Zhu, W., Pan, Z. C., Sui, Q. J., Shang, L. P., and Zhan, J., 2019. "Cross-resistance of the pathogenic fungus *Alternaria alternata* to fungicides with different modes of action." *BMC Microbiol.*, vol. 19, p. 205.

- [21] Luan, F., Wei, L., Zhang, J., Mi, Y., Dong, F., Li, Q., and Guo, Z., 2018. "Antioxidant activity and antifungal activity of chitosan derivatives with propane sulfonate groups." *Polymers (Basel)*, vol. 10, p. E395.
- [22] Hasnaoui, O., Adli, D. E., Halla, N., and Kahloula, K., 2014. "Evaluation de l'activité antifongique des huiles essentielles de *Chamaerops humilis* L. sur des souches isolées des silos de stockage." *PhytoChem and BioSub Journal*, vol. 8, pp. 221-228.
- [23] Sakkas, H. and Papadopoulou, C., 2017. "Antimicrobial activity of basil, oregano, and thyme essential oils." *J. Microbiol Biotechnol.*, vol. 27, pp. 429-438.
- [24] Segvić Klarić, M., Kosalec, I., Mastelić, J., Piecková, E., and Pepeljnak, S., 2007. "Antifungal activity of thyme (*Thymus vulgaris* L.) essential oil and thymol against moulds from damp dwellings." *Lett. Appl. Microbiol.*, vol. 44, pp. 36-42.
- [25] Hadizadeh, I., Peivastegan, B., and Hamzehzarghani, H., 2009. "Antifungal activity of essential oils from some medicinal plants of iran against *alternaria alternata*." *American Journal of Applied Sciences*, vol. 6, pp. 744-748.
- [26] Perina, F. J., Amaral, D. C., Fernandes, R. S., Labory, C. R., Teixeira, G. A., and Alves, E., 2015. "Thymus vulgaris essential oil and thymol against *Alternaria alternata* (Fr.) Keissler: effects on growth, viability, early infection and cellular mode of action." *Pest Manag. Sci.*, vol. 71, pp. 1371-8.
- [27] Puškárová, A., Bučková, M., Kraková, L., Pangallo, D., and Kozics, K., 2017. "The antibacterial and antifungal activity of six essential oils and their cyto/genotoxicity to human HEL 12469 cells." *Sci Rep.*, vol. 7, p. 8211.
- [28] Hong, J. K., Jo, Y. S., Ryoo, D. H., Jung, J. H., Kwon, H. J., Lee, Y. H., Chang, S. W., and Park, C., 2018. "Alternaria spots in tomato leaves differently delayed by four plant essential oil vapours." *Res. Plant Dis.*, vol. 24, pp. 292-301. Available: <https://doi.org/RPD-24-292>
- [29] Fani, M. and Kohanteb, J., 2017. "In vitro antimicrobial activity of thymus vulgaris essential oil against major oral pathogens." *J. Evid. Based Complementary Altern. Med.*, vol. 22, pp. 660-666.
- [30] Gedikoğlu, A., Sökmen, M., and Çivit, A., 2019. "Evaluation of *Thymus vulgaris* and *Thymbra spicata* essential oils and plant extracts for chemical composition, antioxidant, and antimicrobial properties." *Food Sci Nutr.*, vol. 7, pp. 1704-1714.
- [31] Botelho, M. A., Nogueira, N. A. P., Bastos, G. M., Fonseca, S. G. C., Lemos, T. L. G., Matos, F. J. A., Montenegro, D., Heukelbach, J., Rao, V. S., et al., 2007. "Antimicrobial activity of the essential oil from *Lippia sidoides*, carvacrol and thymol against oral pathogens." *Brazilian Journal of Medical and Biological Research*, vol. 40, pp. 349-356.
- [32] Du, E., Gan, L., Li, Z., Wang, W., Liu, D., and Guo, Y., 2015. "In vitro antibacterial activity of thymol and carvacrol and their effects on broiler chickens challenged with *Clostridium perfringens*." *J. Anim. Sci. Biotechnol.*, vol. 6, p. 58.
- [33] Lambert, R., Skandamis, P. N., Coote, P. J., and Nychas, G. J., 2001. "A study of the minimum inhibitory concentration and mode of action of oregano essential oil, thymol and carvacrol." *J. Appl Microbiol.*, vol. 91, pp. 453-462.
- [34] Trombetta, D., Castelli, F., Sarpietro, M. G., Venuti, V., Cristani, M., Daniele, C., Saija, A., Mazzanti, G., and Bisignano, G., 2005. "Mechanisms of antibacterial action of three monoterpenes." *Antimicrob Agents Chemother.*, vol. 49, pp. 2474-2478.
- [35] Lopez-Romero, J. C., Gonzales-Rios, H., Borges, A., and Simoes, M., 2015. "Antibacterial effects and mode of action of selected essential oils components against *Escherichia coli* and *Staphylococcus aureus*." *Evid. Based Complement. Altern. Med.*, vol. 2015, pp. 1-9. Available: <http://dx.doi.org/10.1155/2015/795435>
- [36] Yanishlieva, N. V., Marinova, E. M., Gordon, M. H., and Raneva, V. G., 1999. "Antioxidant activity and mechanism of action of thymol and carvacrol in two lipid systems." *Food Chem.*, vol. 64, pp. 59-66.
- [37] Bennis, S., Chami, F., Chami, N., Bouchikhi, T., and Remmal, A., 2004. "Surface alteration of *Saccharomyces cerevisiae* induced by thymol and eugenol." *Lett. Appl. Microbiol.*, vol. 38, pp. 454-458.
- [38] Vasconcelos, L., Sampaio, F. C., Albuquerque, A., and Vasconcelos, L., 2014. "Cell viability of *Candida albicans* against the antifungal activity of thymol." *Braz. Dent. J.*, vol. 25, pp. 277-281.
- [39] Jassim, N. S., 2017. "Evaluation of some plant extract against the growth of *Alternaria alternata* as one of the causal agent of leaf spots disease of date palm phoenix *dactylifera* L." *Basrah Journal for Date Palm Research*, vol. 16, pp. 75-91.
- [40] Zarai, Z., Ben, C. I., Ben, M. R., Békir, A., Gharsallah, N., and Kadri, A., 2012. "Essential oil of the leaves of *Ricinus communis* L.: in vitro cytotoxicity and antimicrobial properties." *Lipids Health Dis.*, vol. 11, p. 102.
- [41] Mahboubi, M., Kazempour, N., and Valian, M., 2013. "Antimicrobial activity of natural Respitol-B and its main components against poultry microorganisms." *Pak. J. Biol. Sci.*, vol. 16, pp. 1065-8.
- [42] Swamy, M. K., Akhtar, M. S., and Sinniah, U. R., 2016. "Antimicrobial properties of plant essential oils against human pathogens and their mode of action: An updated review, evidence-based complementary and alternative medicine." vol. 2016, p. 21. Available: <https://doi.org/10.1155/2016/3012462>
- [43] Taskeen, U. N., Wani, A. H., and Mir, R. A., 2010. "Antimycotic activity of plant extracts on the spore germination of some pathogenic fungi." *Mycopath.*, vol. 8, pp. 65-69.

- [44] Şesan, T. E., Enache, E., Iacomi, B., Oprea, M., Oancea, F., and Iacomi, C., 2016. "Antifungal activity of some plant extracts against *Alternaria alternata* (Fr.) Keissl. in the blackcurrant crop (*Ribes nigrum* L.)." *Acta Sci. Pol. Hortorum Cultus*, vol. 15, pp. 57–68.
- [45] Alseeni, M. N., Allheani, E. M., Qusti, S. Y., Qusti, N. Y., Alotaibi, S. A., and Alotaibi, H. A., 2019. "Antimicrobial activities of some plant extracts against some fungal pathogens." *IOSR-JPBS*, vol. 14, pp. 01-10.
- [46] Yin, M. C. and Cheng, W. S., 1998. "Inhibition of *Aspergillus niger* and *Aspergillus flavus* by some herbs and spices." *J. Food Prot.*, vol. 61, pp. 123-5.
- [47] Fawzi, E. M., Khalil, A. A., and Afifi, A. F., 2009. "Antifungal effect of some plant extracts on *Alternaria alternata* and *Fusarium oxysporum*." *Afr. J. Biotechnol.*, vol. 8, pp. 2590–2597.
- [48] Sharma, N. and Tiwari, R., 2013. "Biological effects of ginger (*Zingiber officinale* roscoe.) essential oil on *Alternaria alternata* (fr.) keissl." *International Journal of Recent Scientific Research*, vol. 4, pp. 827-831.
- [49] Osman, E. H. B., Algam, S. A. E., Osman, T. M. T., Ali, M. E., Abbo, A. S. H., and Elhassan, S. M., 2016. "Antifungal effect of camel urine and ginger water extract against *Alternaria alternata* the causal agent of early blight disease of tomato in vitro." *International Journal of Agriculture, Forestry and Plantation*, vol. 2, pp. 261-69.
- [50] Ahmad, M. S. and Qureshi, A. H., 2017. "Antifungal activity of *Zingiber officinale* oil against plant pathogenic fungi isolated from solanaceous vegetable fruits." *Asian Journal of Pharmacy and Pharmacology*, vol. 3, pp. 121-124.
- [51] Masniari, P., 2011. "The effect of red ginger (*Zingiber officinale* Roscoe) extract on the growth of mastitis causing bacterial isolates." *African Journal of Microbiology Research*, vol. 5, pp. 382-389.
- [52] Ali, B., Al-Wabel, N. A., Shams, S., Ahamad, A., Khan, S. A., and Anwar, F., 2015. "Essential oils used in aromatherapy: a systemic review." *Asian Pacific Journal of Tropical Biomedicine*, vol. 5, pp. 601–611.
- [53] Fei, L., Fei, G., Qianqian, W., and Lin, L., 2017. "Changes of membrane fatty acids and proteins of *Shewanella putrefaciens* treated with cinnamon oil and gamma irradiation." *Bioresources and Bioprocessing*, vol. 4, p. 1.
- [54] Kloucek, P., Smid, J., Frankova, A., Kokoska, L., Valterova, I., and Pavela, R., 2012. "Fast screening method for assessment of antimicrobial activity of essential oils in vapor phase." *Food Res. Int.*, vol. 47, pp. 161-165.
- [55] Rizwana, H., Alwhibi, M. S., and Soliman, D. A., 2016. "Antimicrobial activity and chemical composition of flowers of *matricaria aurea* a native herb of Saudi Arabia." *International Journal of Pharmacology*, vol. 12, pp. 576-586.
- [56] Al-Snafi, A. E., 2016. "Medical importance of *Anthemis nobilis* (*Chamaemelum nobile*) - a review." *Asian Journal of Pharmaceutical Science and Technology*, vol. 6, pp. 89-95.
- [57] Móricz, A. M., Szarka, S., Ott, P. G., Héthelyi, E. B., Szoke, E., and Tyihák, E., 2012. "Separation and identification of antibacterial chamomile components using OPLC, bioautography and GC-MS." *Med. Chem.*, vol. 8, pp. 85-94.
- [58] Al-Kuraishy, H. M., Al-Gareeb, I., Albuhadilly, K. A., and Alwindy, S., 2015. "In vitro assessment of the antibacterial activity of *Matricaria chamomile* alcoholic extract against pathogenic bacterial strains." *Br. Microbiol. Res. J.*, vol. 7, pp. 55–61.
- [59] Mahmoud, A. M., Abd El-Baky, R. M., Ahmed, A. F., and Mahmoud, G. G. F., 2016. "Antibacterial activity of essential oils and in combination with some standard antimicrobials against different pathogens isolated from some clinical specimens." *American Journal of Microbiological Research*, vol. 4, pp. 16-25.
- [60] Aggag, M. E. and Yousef, R. T., 1972. "Study of antimicrobial activity of chamomile oil." *Planta Med.*, vol. 22, pp. 140-144.
- [61] Kedzia, B., 1991. "Antimicrobial activity of chamomile and its components." *Herba Polonica*, vol. 37, pp. 29-38.
- [62] Lu, T., Cantrell, C. L., Robbs, S. L., Franzblau, S. G., and Fischer, N. H., 1998. "Antimycobacterial *matricaria* esters and lactones from *astereae* species." *Planta Medica*, vol. 64, pp. 665-667.
- [63] Szalontai, M., Verzar, P. O., and Florian, E., 1976. "Data on the antifungal effect of the biologically active components of *Matricaria chamomilla* L." *Acta Pharm. Hung.*, vol. 46, pp. 232-247.
- [64] Szalontai, M., Petri-Verzar, G., and Florian, E., 1977. "Contribution to the study of antimycotic effect of biologically active components of *Matricaria chamomilla* L." *Perfumery Cosmet.*, vol. 58, pp. 121-127.
- [65] Ahmed, F. H., El Badri, A. A., Ibrahim, M. M. K., El Shahed, A. S., and El Khalafawy, H. M. M., 1994. "Comparative studies of antifungal potentialities for some natural plant oils against different fungi isolated from poultry." *Fats Oils*, vol. 45, pp. 260-264.